

Резюме: Лебедев Юрий Борисович

Адрес

Федеральное государственное бюджетное учреждение науки Институт биоорганической химии им. академиков М.М. Шемякина и Ю.А. Овчинникова Российской академии наук, Москва, Россия

Контакты

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Образование

1973–1978

Москва

МГУ

Работа в ИБХ

2018–наст.вр.

Главный научный сотрудник

Членство в советах и комиссиях ИБХ

Диссертационный совет

Ученый совет

Научные интересы

Руководитель лаборатории доктор биологических наук Юрий Борисович Лебедев является одним из ведущих отечественных специалистов в области изучения структуры, функционирования и эволюции генома млекопитающих. Исследования в этой области начаты Ю. Б. Лебедевым в начале 1990-х, когда он с группой молодых ученых лаборатории академика Е. Д. Свердлова вошли в число первых участников Российской программы «Геном Человека». Как руководитель лаборатории Ю. Б. Лебедев продолжает разрабатывать оригинальное направление эволюционной геномики, связанное с исследованием функциональных последствий распространения и активации ретропозонов в геномах высших приматов.

Членство в сообществах

В 1996 г. Лебедев Ю. Б. был принят в члены HUGO.

Степени и звания

Профессор

Доктор наук (Биологические науки, 03.00.03 — Молекулярная биология)

Гранты и проекты

2020–2022 [Взаимодействие генотипа и репертуара Т-лимфоцитов при формировании противовирусного иммунного ответа](#)

2015–2019 [Изучение динамики системы клеточного адаптивного иммунитета при флавивирусной инфекции](#)

2017–2020 [Анализ клонального спектра малых субпопуляций Т-лимфоцитов при формировании иммунного ответа у человека](#)

Публикации

1. Smirnova AO, Miroshnichenkova AM, Olshanskaya YV, Maschan MA, **Lebedev YB**, Chudakov DM, Mamedov IZ, Komkov A (2023). The use of non-functional clonotypes as a natural calibrator for quantitative bias correction in adaptive immune receptor repertoire profiling. *Elife* 12, , [10.7554/eLife.69157](https://doi.org/10.7554/eLife.69157)
2. Komech EA, Koltakova AD, Barinova AA, Minervina AA, Salnikova MA, Shmidt EI, Korotaeva TV, Loginova EY, Erdes SF, Bogdanova EA, Shugay M, Lukyanov S, **Lebedev YB**, Zvyagin IV (2022). TCR repertoire profiling revealed antigen-driven CD8+ T cell clonal groups shared in synovial fluid of patients with spondyloarthritis. *Front Immunol* 13, 973243, [10.3389/fimmu.2022.973243](https://doi.org/10.3389/fimmu.2022.973243)
3. Sycheva AL, Komech EA, Pogorelyy MV, Minervina AA, Urazbakhtin SZ, Salnikova MA, Vorovitch MF, Kopantzev EP, Zvyagin IV, Komkov AY, Mamedov IZ, **Lebedev YB** (2022). Inactivated tick-borne encephalitis vaccine elicits several overlapping waves of T cell response. *Front Immunol* 13, 970285, [10.3389/fimmu.2022.970285](https://doi.org/10.3389/fimmu.2022.970285)
4. Urazbakhtin S, Smirnova A, Volakhava A, Zerkalenkova E, Salyutina M, Doubek M, Jelinkova H, Khudainazarova N, Volchkov E, Belyaeva L, Komech E, Pavlova S, **Lebedev Y**, Plevova K, Olshanskaya Y, Komkov A, Mamedov I (2022). The Absence of Retroelement Activity Is Characteristic for Childhood Acute Leukemias and Adult Acute Lymphoblastic Leukemia. *Int J Mol Sci* 23 (3), , [10.3390/ijms23031756](https://doi.org/10.3390/ijms23031756)
5. **(конференция)** Sycheva AL, Pogorelyy MV, Komech EA, Urazbakhtin SZ, Minervina AA, Kopancev EP, Vorovitch MF, Zvyagin IV, Mamedov IZ, **Lebedev YB** (2021). Features of T-cell immune response to tick-borne encephalitis vaccine. *Eur J Immunol* 51 (S1), 1–448, <https://doi.org/10.1002/eji.202170200>
6. **(конференция)** Zvyagin IV, Blagov S, Fomchenkova V, Fadeeva M, Komech EA, Zhogov V, Barinova AA, Mikelov AI, Sycheva AL, **Lebedev YB**, Maschan MA (2021). T cell repertoire sequencing to study the contribution of different donor T cell subsets to patient repertoire at the early stage after $\alpha\beta$ T/CD19-depleted allogeneic hematopoietic stem cell transplantation. *Eur J Immunol* 51 (S1), 1–448, <https://doi.org/10.1002/eji.202170200>
7. **(конференция)** Комеч ЕА, Звягин ИВ, **Лебедев ЮБ**, Сальникова МА, Минервина АА (2021). T-CELL REPERTOIRE OF SYNOVIAL FLUID IN SPONDYLOARTHROPATHIES EXHIBITS HALLMARKS OF HLA-DEPENDENT CLONAL EXPANSIONS AND REMAINS STABLE OVER 1.5 YEARS. *Ann Rheum Dis* (80), 204, [10.1136/annrheumdis-2021-eular.3498](https://doi.org/10.1136/annrheumdis-2021-eular.3498)
8. Kovalenko EI, Zvyagin IV, Streltsova MA, Mikelov AI, Erokhina SA, Telford G, Sapozhnikov AM, **Lebedev YB** (2021). Surface NKG2C identifies differentiated $\alpha\beta$ T-cell clones expanded in peripheral blood. *Front Immunol* 11, 613882, [10.3389/fimmu.2020.613882](https://doi.org/10.3389/fimmu.2020.613882)
9. Minervina AA, Komech EA, Titov A, Koraiichi MB, Rosati E, Mamedov IZ, Franke A, Efimov GA, Chudakov DM, Mora T, Walczak AM, **Lebedev YB**, Pogorelyy MV (2021). Longitudinal high-throughput TCR repertoire profiling reveals the dynamics of T-cell memory formation after mild COVID-19 infection. *Elife* 10, 1–17, [10.7554/eLife.63502](https://doi.org/10.7554/eLife.63502)
10. Komkov AY, Urazbakhtin SZ, Saliutina MV, Komech EA, Shelygin YA, Nugmanov GA, Shubin VP, Smirnova AO, Bobrov MY, Tsukanov AS, Snezhkina AV, Kudryavtseva AV, **Lebedev YB**, Mamedov IZ (2020). SeqURE – a new copy-capture based method for sequencing of unknown Retroinsertion events. *Mob DNA* 11 (1), 33, [10.1186/s13100-020-00228-6](https://doi.org/10.1186/s13100-020-00228-6)
11. **(конференция)** Mikelov AI, Komech EA, **Lebedev YB**, Zvyagin IV (2020). In- and off- season peripheral blood T cell repertoire profiling of patients with birch pollen allergy. *Allergy* 75 (S109), 188, [10.1111/all.14506](https://doi.org/10.1111/all.14506)
12. **(конференция)** Комков АЮ, Мамедов ИЗ, **Лебедев ЮБ**, Атапина Е (2020). A cost-effective quasi single-cell assay for deciphering of clonal architecture of leukemic cells. *Klin Padiatr* 232 (3), e7, [10.1055/s-0040-1709799](https://doi.org/10.1055/s-0040-1709799)
13. Minervina AA, Pogorelyy MV, Komech EA, Karnaukhov VK, Bacher P, Rosati E, Franke A, Chudakov D, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2020). Primary and secondary anti-viral response captured by the dynamics and phenotype of individual T cell clones. *Elife* 9, , [10.7554/eLife.53704](https://doi.org/10.7554/eLife.53704)
14. Rosati E, Pogorelyy MV, Dowds CM, Moller FT, Sorensen SB, **Lebedev YB**, Frey N, Schreiber S, Spehlmann ME, Andersen V, Mamedov IZ, Franke A (2019). Identification of disease-associated traits and clonotypes in the T-cell receptor repertoire of monozygotic twins affected by inflammatory bowel diseases. *J Crohns Colitis* 14 (6), 778–790, [10.1093/ecco-jcc/jjz179](https://doi.org/10.1093/ecco-jcc/jjz179)
15. Микелов АИ, Староверов ДБ, Комеч ЕА, **Лебедев ЮБ**, Чудаков ДМ, Zvyagin IV (2019). Correlated dynamics of serum IGE and IGE+ clonotype count with allergen air level in seasonal allergic rhinitis. *Bulletin*

- of Russian State Medical University 5 (5), 13–22, [10.24075/brsmu.2019.072](https://doi.org/10.24075/brsmu.2019.072)
16. Komkov A, Miroshnichenkova A, Nugmanov G, Popov A, Pogorelyy M, Zapletalova E, Jelinkova H, Pospisilova S, **Lebedev Y**, Chudakov D, Olshanskaya Y, Plevova K, Maschan M, Mamedov I (2019). High-throughput sequencing of T-cell receptor alpha chain clonal rearrangements at the DNA level in lymphoid malignancies. *Br J Haematol* 188 (5), 723–731, [10.1111/bjh.16230](https://doi.org/10.1111/bjh.16230)
 17. **(конференция)** Mikelov AI, Turchaninova MA, Komech EA, Staroverov DB, Shvets SM, **Lebedev YB**, Chudakov DM, Zvyagin IV (2019). Longitudinal profiling of immunoglobulin heavy-chain repertoires in memory B-cells, plasmablasts and plasma cells from peripheral blood of individuals with birch pollen allergy. *Allergy* 74 (S106), 174.
 18. Pogorelyy MV, Minervina AA, Shugay M, Chudakov DM, **Lebedev YB**, Mora T, Walczak AM (2019). Detecting T cell receptors involved in immune responses from single repertoire snapshots. *PLoS Biol* 17 (6), e3000314, [10.1371/journal.pbio.3000314](https://doi.org/10.1371/journal.pbio.3000314)
 19. Nugmanov GA, Komkov AY, Saliutina MV, Minervina AA, **Lebedev YB**, Mamedov IZ (2019). [A Pipeline for the Error-free Identification of Somatic Alu Insertions in High-throughput Sequencing Data]. *Mol Biol (Mosk)* 53 (1), 154–165, [10.1134/S0026898419010117](https://doi.org/10.1134/S0026898419010117)
 20. Nugmanov GA, Komkov AY, Saliutina MV, Minervina AA, **Lebedev YB**, Mamedov IZ (2019). A Pipeline for the Error-Free Identification of Somatic Alu Insertions in High-Throughput Sequencing Data. *Mol Biol* 53 (1), 138–146, [10.1134/S0026893319010114](https://doi.org/10.1134/S0026893319010114)
 21. Pogorelyy MV, Minervina AA, Touzel MP, Sycheva AL, Komech EA, Kovalenko EI, Karganova GG, Egorov ES, Komkov AY, Chudakov DM, Mamedov IZ, Mora T, Walczak AM, **Lebedev YB** (2018). Precise tracking of vaccine-responding T cell clones reveals convergent and personalized response in identical twins. *Proc Natl Acad Sci U S A* 115 (50), 12704–12709, [10.1073/pnas.1809642115](https://doi.org/10.1073/pnas.1809642115)
 22. Komkov AY, Minervina AA, Nugmanov GA, Saliutina MV, **Lebedev YB**, Mamedov IZ, Khodosevich KV (2018). An advanced enrichment method for rare somatic retroelement insertions sequencing. *Mob DNA* 9 (1), 31, [10.1186/s13100-018-0136-1](https://doi.org/10.1186/s13100-018-0136-1)
 23. **(конференция)** Комеч ЕА, Колтакова АД, Мясоутова АА, Коротаева ТВ, Шмидт НИ, Шостак НА, **Лебедев ЮБ**, Звягин ИВ (2018). Клональная характеристика Т-лимфоцитов очага воспаления у больных со спондилоартропатиями. *Nauchno-Prakticheskaya Revmatologiya* 56 (3), 91.
 24. **(конференция)** Звягин ИВ, **Лебедев ЮБ**, Мясоутова АА, Комеч ЕА (2018). TCRbeta CDR3 motif is detected in synovial fluid of patients with different spondyloarthropathies. *FEBS Open Bio* 8, 489.
 25. **(конференция)** Fomchenkova E, Komech A, Blagov , Sycheva L, **Lebedev B**, Chudakov M, Maschan A, Zvyagin V (2018). T cell repertoire profiling after hematopoietic stem cell transplantation with CD19/αβT cell depletion and donor lymphocyte infusion. *FEBS Open Bio* 8 (S1), 281: P.09–230–Tue.
 26. Komech EA, Pogorelyy MV, Egorov ES, Britanova OV, Rebrikov DV, Bochkova AG, Shmidt EI, Shostak NA, Shugay M, Lukyanov S, Mamedov IZ, **Lebedev YB**, Chudakov DM, Zvyagin IV (2018). CD8+T cells with characteristic T cell receptor beta motif are detected in blood and expanded in synovial fluid of ankylosing spondylitis patients. *Rheumatology (Oxford)* 57 (6), 1097–1104, [10.1093/rheumatology/kex517](https://doi.org/10.1093/rheumatology/kex517)
 27. Komech EA, Zvyagin IV, Pogorelyy MV, Mamedov IZ, Fedorenko DA, **Lebedev YB** (2018). Characterization of the T-cell repertoire after autologous HSCT in patients with ankylosing spondylitis. *Acta Naturae* 10 (2), 48–57, [10.32607/2075851-2018-10-2-48-57](https://doi.org/10.32607/2075851-2018-10-2-48-57)
 28. Sycheva AL, Pogorelyy MV, Komech EA, Minervina AA, Zvyagin IV, Staroverov DB, Chudakov DM, **Lebedev YB**, Mamedov IZ (2018). Quantitative profiling reveals minor changes of T cell receptor repertoire in response to subunit inactivated influenza vaccine. *Vaccine* 36 (12), 1599–1605, [10.1016/j.vaccine.2018.02.027](https://doi.org/10.1016/j.vaccine.2018.02.027)
 29. Pogorelyy MV, Minervina AA, Chudakov DM, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2018). Method for identification of condition-associated public antigen receptor sequences. *Elife* 7, , [10.7554/eLife.33050](https://doi.org/10.7554/eLife.33050)
 30. Komech EA, **Lebedev YB**, Koshenkova AV, Syrko DS, Musatkina EA, Lukyanov SA, Chudakov DM, Zvyagin IV (2018). A study of the repertoire of activated T-cell clones obtained from a patient with ankylosing spondylitis. *Bulletin of Russian State Medical University* 7 (1), 65–73, [10.24075/brsmu.2018.001](https://doi.org/10.24075/brsmu.2018.001)
 31. Komech EA, Zvyagin IV, Pogorelyy MV, Mamedov IZ, Fedorenko DA, **Lebedev YB** (2018). Characterization of the T-cell Repertoire after Autologous HSCT in Patients with Ankylosing Spondylitis. *Acta Naturae* 10 (2), 48–57, [10.32607/20758251-2018-10-2-48-57](https://doi.org/10.32607/20758251-2018-10-2-48-57)

32. Pogorelyy MV, Elhanati Y, Marcou Q, Sycheva AL, Komech EA, Nazarov VI, Britanova OV, Chudakov DM, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2017). Persisting fetal clonotypes influence the structure and overlap of adult human T cell receptor repertoires. *PLoS Comput Biol* 13 (7), e1005572, [10.1371/journal.pcbi.1005572](https://doi.org/10.1371/journal.pcbi.1005572)
33. Komkov A, Miroshnichenkova A, Minervina A, Nugmanov G, **Lebedev Y**, Mamedov I, Olshanskaya Y, Maschan M (2017). High-throughput sequencing for diagnostics of minimal residual disease in acute lymphoblastic leukemia. *Klin Padiatr* , , [10.1055/s-0037-1602224](https://doi.org/10.1055/s-0037-1602224)
34. **(конференция)** Pogorelyy M, PuelmaTouzer M, Minervina AA, Sycheva AL, Chudakov DM, Mamedov IZ, Mora T, Walczak AM, **Lebedev YB** (2017). High throughput sequencing of identical twins TCR repertoires after yellow fever vaccination. , 60.
35. Zvyagin IV, Mamedov IZ, Tatarinova OV, Komech EA, Kurnikova EE, Boyakova EV, Brilliantova V, Shelikhova LN, Balashov DN, Shugay M, Sycheva AL, Kasatskaya SA, **Lebedev YB**, Maschan AA, Maschan MA, Chudakov DM (2017). Tracking T-cell immune reconstitution after TCR $\alpha\beta$ /CD19-depleted hematopoietic cells transplantation in children. *Leukemia* 31 (5), 1145–1153, [10.1038/leu.2016.321](https://doi.org/10.1038/leu.2016.321)
36. **(конференция)** Сычева АЛ, Погорельый МВ, Комеч ЕА, Звягин ИВ, Мамедов ИЗ, **Лебедев ЮБ** (2017). Изучение малых субпопуляций активированных Т-лимфоцитов из крови доноров, вакцинированных против вируса жёлтой лихорадки. , 98.
37. **(конференция)** Zvyagin I, Tatarinova O, Mamedov I, Komech E, Maschan A, Shelikhova L, Kurnikova E, Boyakova E, **Lebedev Y**, Maschan M, Chudakov D (2016). T Cell Repertoire after Alpha/Beta-T Cell Depleted Allogeneic Hematopoietic Stem Cell Transplantation in Pediatric Patients. *Blood* (128), 4582.
38. Nazarov VI, Minervina AA, Komkov AY, Pogorelyy MV, Maschan MA, Olshanskaya YV, Zvyagin IV, Chudakov DM, **Lebedev YB**, Mamedov IZ (2016). Reliability of immune receptor rearrangements as genetic markers for minimal residual disease monitoring. *Bone Marrow Transplant* 51 (10), 1408–1410, [10.1038/bmt.2016.148](https://doi.org/10.1038/bmt.2016.148)
39. **(конференция)** Komkov AY, Minervina AA, Pogorelyy MV, Zvyagin IV, Panferova A, Olshanskaya Y, Chudakov DM, Maschan M, Mamedov IZ, **Lebedev YB** (2016). Next generation sequencing based approach for monitoring of minimal residual disease in acute lymphoblastic leukemia. *FEBS J* 283 (S1), 376, [10.1111/febs.13808](https://doi.org/10.1111/febs.13808)
40. **(конференция)** Погорельый МВ, Сычева АЛ, Мамедов ИЗ, Мора Т, Вальзак АМ, **Лебедев ЮБ** (2016). Клоны Т-клеток пуповинной крови обнаруживаются в репертуарах Т-клеточных рецепторов взрослых доноров. , 23.
41. Minervina AA, Komkov AY, Mamedov IZ, **Lebedev YB** (2016). Advanced lymphoblastic clones detection in T-cell leukemia. *Dokl Biochem Biophys* 467 (1), 85–88, [10.1134/S1607672916020022](https://doi.org/10.1134/S1607672916020022)
42. **(конференция)** Сычева АЛ, Погорельый МВ, Комеч ЕА, Мамедов ИЗ, **Лебедев ЮБ** (2016). Динамика Т-клеточного репертуара человека в ходе противогриппозной вакцинации и ревакцинации. , 27.
43. Komkov AY, Miroshnichenkova AM, Olshanskaya YV, Myakova NV, Diakonova YY, Minervina AL, Mamedov IZ, **Lebedev YB**, Maschan AA, Maschan MA (2016). Detection of immunoglobulin genes rearrangements in patients with acute lymphoblastic leukemia using highthroughput next generation sequencing. *Probl Gematol Pereliv Krovi* 61 (4), 200–204, [10.18821/0234-5730-2016-61-4-200-204](https://doi.org/10.18821/0234-5730-2016-61-4-200-204)
44. Nazarov VI, Pogorelyy MV, Komech EA, Zvyagin IV, Bolotin DA, Shugay M, Chudakov DM, **Lebedev YB**, Mamedov IZ (2015). tcR: An R package for T cell receptor repertoire advanced data analysis. *BMC Bioinformatics* 16 (1), 175, [10.1186/s12859-015-0613-1](https://doi.org/10.1186/s12859-015-0613-1)
45. **(конференция)** Pogorelyy MV, Sycheva AL, Komech EA, Marcou Q, Elhanati Y, Mora T, Walczak A, Mamedov IZ, **Lebedev YB** (2015). Deep TCR repertoire profiling after seasonal influenza vaccination. , 432.
46. Kurnosov AA, Ustyugova SV, Nazarov VI, Minervina AA, Komkov AY, Shugay M, Pogorelyy MV, Khodosevich KV, Mamedov IZ, **Lebedev YB** (2015). The evidence for increased L1 activity in the site of human adult brain neurogenesis. *PLoS One* 10 (2), e0117854, [10.1371/journal.pone.0117854](https://doi.org/10.1371/journal.pone.0117854)
47. Egorov ES, Merzlyak EM, Shelenkov AA, Britanova OV, Sharonov GV, Staroverov DB, Bolotin DA, Davydov AN, Barsova E, **Lebedev YB**, Shugay M, Chudakov DM (2015). Quantitative profiling of immune repertoires for minor lymphocyte counts using unique molecular identifiers. *J Immunol* 194 (12), 6155–6163, [10.4049/jimmunol.1500215](https://doi.org/10.4049/jimmunol.1500215)
48. Zvyagin IV, Pogorelyy MV, Ivanova ME, Komech EA, Shugay M, Bolotin DA, Shelenkov AA, Kurnosov AA,

- Staroverov DB, Chudakov DM, **Lebedev YB**, Mamedov IZ (2014). Distinctive properties of identical twins' TCR repertoires revealed by high-throughput sequencing. *Proc Natl Acad Sci U S A* 111 (16), 5980–5985, [10.1073/pnas.1319389111](https://doi.org/10.1073/pnas.1319389111)
49. Britanova OV, Putintseva EV, Shugay M, Merzlyak EM, Turchaninova MA, Staroverov DB, Bolotin DA, Lukyanov S, Bogdanova EA, Mamedov IZ, **Lebedev YB**, Chudakov DM (2014). Age-Related decrease in TCR repertoire diversity measured with deep and normalized sequence profiling. *J Immunol* 192 (6), 2689–2698, [10.4049/jimmunol.1302064](https://doi.org/10.4049/jimmunol.1302064)
50. Mamedov IZ, Britanova OV, Zvyagin IV, Turchaninova MA, Bolotin DA, Putintseva EV, **Lebedev YB**, Chudakov DM (2013). Preparing unbiased T-cell receptor and antibody cDNA libraries for the deep next generation sequencing profiling. *Front Immunol* 4 (DEC), 456, [10.3389/fimmu.2013.00456](https://doi.org/10.3389/fimmu.2013.00456)
51. Putintseva EV, Britanova OV, Staroverov DB, Merzlyak EM, Turchaninova MA, Shugay M, Bolotin DA, Pogorelyy MV, Mamedov IZ, Bobrynina V, Maschan M, **Lebedev YB**, Chudakov DM (2013). Mother and child T cell receptor repertoires: Deep profiling study. *Front Immunol* 4 (DEC), 463, [10.3389/fimmu.2013.00463](https://doi.org/10.3389/fimmu.2013.00463)
52. Kurnosov AA, Ustyugova SV, Pogorelyy MV, Komkov AY, Bolotin DA, Khodosevich KV, Mamedov IZ, **Lebedev YB** (2013). A novel approach to identification of somatic retroelements' insertions in human genome. *Russ. J. Bioorganic Chem.* 39 (4), 417–425, [10.1134/S1068162013040110](https://doi.org/10.1134/S1068162013040110)
53. Bolotin DA, Mamedov IZ, Britanova OV, Zvyagin IV, Shagin D, Ustyugova SV, Turchaninova MA, Lukyanov S, **Lebedev YB**, Chudakov DM (2012). Next generation sequencing for TCR repertoire profiling: Platform-specific features and correction algorithms. *Eur J Immunol* 42 (11), 3073–3083, [10.1002/eji.201242517](https://doi.org/10.1002/eji.201242517)
54. Britanova OV, Bochkova AG, Staroverov DB, Fedorenko DA, Bolotin DA, Mamedov IZ, Turchaninova MA, Putintseva EV, Kotlobay AA, Lukyanov S, Novik AA, **Lebedev YB**, Chudakov DM (2012). First autologous hematopoietic SCT for ankylosing spondylitis: A case report and clues to understanding the therapy. *Bone Marrow Transplant* 47 (11), 1479–1481, [10.1038/bmt.2012.44](https://doi.org/10.1038/bmt.2012.44)
55. Komkov AY, Maschan MA, Shvets VI, **Lebedev YB** (2012). Functional analysis of polymorphic insertions of alu retroelements in acute lymphoblastic leukemia patients. *Russ. J. Bioorganic Chem.* 38 (3), 306–318, [10.1134/S1068162012030089](https://doi.org/10.1134/S1068162012030089)
56. Mamedov IZ, Ustyugova SV, Amosova FL, **Lebedev YB** (2011). Retroelement insertion polymorphism and modulation of human gene activity. , 203–236.
57. Mamedov IZ, Britanova OV, Bolotin DA, Chkalina AV, Staroverov DB, Zvyagin IV, Kotlobay AA, Turchaninova MA, Fedorenko DA, Novik AA, Sharonov GV, Lukyanov S, Chudakov DM, **Lebedev YB** (2011). Quantitative tracking of T cell clones after haematopoietic stem cell transplantation. *EMBO Mol Med* 3 (4), 201–207, [10.1002/emmm.201100129](https://doi.org/10.1002/emmm.201100129)
58. Zvyagin IV, Mamedov IZ, Britanova OV, Staroverov DB, Nasonov EL, Bochkova AG, Chkalina AV, Kotlobay AA, Korostin DO, Rebrikov DV, Lukyanov S, **Lebedev YB**, Chudakov DM (2010). Contribution of functional KIR3DL1 to ankylosing spondylitis. *Cell Mol Immunol* 7 (6), 471–476, [10.1038/cmi.2010.42](https://doi.org/10.1038/cmi.2010.42)
59. Zvyagin IV, Dorodnykh VY, Mamedov IZ, Staroverov DB, Bochkova AG, Rebrikov DV, **Lebedev YB** (2010). Association of ERAP1 Allelic Variants with Risk of Ankylosing Spondylitis. *Acta Naturae* 2 (3), 72–7.
60. Mamedov IZ, Shagina IA, Kurnikova MA, Novozhilov SN, Shagin DA, **Lebedev YB** (2010). A new set of markers for human identification based on 32 polymorphic Alu insertions. *Eur J Hum Genet* 18 (7), 808–814, [10.1038/ejhg.2010.22](https://doi.org/10.1038/ejhg.2010.22)
61. Chkalina AV, Zvyagin IV, Mamedov IZ, Britanova OV, Staroverov DB, **Lebedev YB** (2010). The oligoclonal expansion of T cells: The investigation of its stability over time. *Russ. J. Bioorganic Chem.* 36 (2), 191–198, [10.1134/S1068162010020081](https://doi.org/10.1134/S1068162010020081)
62. Shagina I, Bogdanova E, Mamedov IZ, **Lebedev Y**, Lukyanov S, Shagin D (2010). Normalization of genomic DNA using duplex-specific nuclease. *Biotechniques* 48 (6), 455–459, [10.2144/000113422](https://doi.org/10.2144/000113422)
63. Amosova AL, Komkov AY, Ustyugova SV, Mamedov IZ, **Lebedev YB** (2009). Retroposons in modern human genome evolution. *Russ. J. Bioorganic Chem.* 35 (6), 702–710, [10.1134/S1068162009060053](https://doi.org/10.1134/S1068162009060053)
64. Mamedov IZ, Britanova OV, Chkalina AV, Staroverov DB, Amosova AL, Mishin AS, Kurnikova MA, Zvyagin IV, Mutovina ZY, Gordeev AV, Khaidukov SV, Sharonov GV, Shagin DA, Chudakov DM, **Lebedev YB** (2009). Individual characterization of stably expanded T cell clones in ankylosing spondylitis patients. *Autoimmunity* 42 (6), 525–536, [10.1080/08916930902960362](https://doi.org/10.1080/08916930902960362)
65. Panchin AY, Spirin SA, Lukyanov SA, **Lebedev YB**, Panchin YV (2008). Human trash ESTs - Sequences

- from cDNA collection that are not aligned to genome assembly. *J Bioinform Comput Biol* 6 (4), 759–773, [10.1142/S0219720008003709](https://doi.org/10.1142/S0219720008003709)
66. Mamedov IZ, Amosova AL, Fisunov GY, **Lebedev YB** (2008). A new polymorphic retroelement database (PRED) for the human genome. *Mol Biol* 42 (4), 641–646, [10.1134/S0026893308040213](https://doi.org/10.1134/S0026893308040213)
 67. **Lebedev YB**, Amosova AL, Mamedov IZ, Fisunov GY, Sverdlov ED (2007). Most recent AluY insertions in human gene introns reduce the content of the primary transcripts in a cell type specific manner. *Gene* 390 (12), 122–129, [10.1016/j.gene.2006.09.031](https://doi.org/10.1016/j.gene.2006.09.031)
 68. Ustyugova SV, **Lebedev YB**, Sverdlov ED (2006). Long L1 insertions in human gene introns specifically reduce the content of corresponding primary transcripts. *Genetica* 128 (13), 261–272, [10.1007/s10709-005-5967-2](https://doi.org/10.1007/s10709-005-5967-2)
 69. Kutuev I, Khusainova R, Karunas A, Yunusbayev B, Fedorova S, **Lebedev Y**, Hunsmann G, Khusnutdinova E (2006). From east to west: Patterns of genetic diversity of populations living in four Eurasian regions. *Hum Hered* 61 (1), 1–9, [10.1159/000091309](https://doi.org/10.1159/000091309)
 70. Ustyugova SV, Amosova AL, **Lebedev YB**, Sverdlov ED (2006). A tissue-specific decrease in the pre-mRNA level of L1- and Alu-containing alleles of human genes. *Russ. J. Bioorganic Chem.* 32 (1), 93–95, [10.1134/S1068162006010110](https://doi.org/10.1134/S1068162006010110)
 71. Buzdin A, Vinogradova T, **Lebedev Y**, Sverdlov E (2005). Genome-wide experimental identification and functional analysis of human specific retroelements. *Cytogenet Genome Res* 110 (14), 468–474, [10.1159/000084980](https://doi.org/10.1159/000084980)
 72. Mamedov IZ, Arzumanyan ES, Amosova AL, **Lebedev YB**, Sverdlov ED (2005). Whole-genome experimental identification of insertion/deletion polymorphisms of interspersed repeats by a new general approach. *Nucleic Acids Res* 33 (2), e16, [10.1093/nar/gni018](https://doi.org/10.1093/nar/gni018)
 73. Ustyugova SV, Amosova AL, **Lebedev YB**, Sverdlov ED (2005). Cell line fingerprinting using retroelement insertion polymorphism. *Biotechniques* 38 (4), 561–565, [10.2144/05384ST02](https://doi.org/10.2144/05384ST02)
 74. Khusainova RI, Akhmetova VL, Kutuyev IA, Salimova AZ, Korshunova TY, **Lebedev YB**, Khusnutdinova EK (2004). Genetic structure of the volga-ural and Central Asian populations inferred from the data on Alu polymorphism. *Genetika* 40 (4), 552–559.
 75. Khodosevich KV, **Lebedev YB**, Sverdlov ED (2004). The tissue-specific methylation of human-specific endogenous retroviral LTRs. *Russ. J. Bioorganic Chem.* 30 (5), 441–445, [10.1023/B:RUJBI.0000043787.07628.2a](https://doi.org/10.1023/B:RUJBI.0000043787.07628.2a)
 76. Mamedov I, **Lebedev Y**, Hunsmann G, Khusnutdinova E, Sverdlov E (2004). A rare event of insertion polymorphism of a HERV-K LTR in the human genome. *Genomics* 84 (3), 596–599, [10.1016/j.ygeno.2004.04.010](https://doi.org/10.1016/j.ygeno.2004.04.010)
 77. Mamedov IZ, **Lebedev YB**, Sverdlov ED (2004). Unusually long target site duplications flanking some of the long terminal repeats of human endogenous retrovirus K in the human genome. *J Gen Virol* 85 (6), 1485–1488, [10.1099/vir.0.19717-0](https://doi.org/10.1099/vir.0.19717-0)
 78. Khusainova RI, Akhmetova VL, Kutuyev IA, Salimova AZ, Korshunova TY, **Lebedev YB**, Khusnutdinova EK (2004). Genetic structure of the Volga-Ural and Central Asian populations inferred from the data on Alu polymorphism. *Russ J Genet* 40 (4), 443–449, [10.1023/B:RUJGE.0000024983.75183.8b](https://doi.org/10.1023/B:RUJGE.0000024983.75183.8b)
 79. Khodosevich K, **Lebedev Y**, Sverdlov ED (2004). Large-scale determination of the methylation status of retrotransposons in different tissues using a methylation tags approach. *Nucleic Acids Res* 32 (3), e31, [10.1093/nar/gnh035](https://doi.org/10.1093/nar/gnh035)
 80. Glazkova DV, Nadezhdin EV, Vinogradova TV, **Lebedev YB**, Bronholdt D, Grzeschik KX, Arman IP, Sverdlov ED (2003). Nucleotide sequences of long terminal repeats of the human endogenous retrovirus (LTR HERV-K) on the short arm of chromosome 7: Identification, analysis, and evaluation of transcriptional activity. *Genetika* 39 (5), 702–708.
 81. Glazkova DV, Nadezhdin EV, Vinogradova TV, **Lebedev YB**, Bronholdt D, Grzeschik KX, Arman IP, Sverdlov ED (2003). Nucleotide sequences of long terminal repeats of the human endogenous retroviruses (LTR HERV-K) on the short arm of chromosome 7: Identification, map locations, and transcriptional activity. *Russ J Genet* 39 (5), 578–583, [10.1023/A:1023744019585](https://doi.org/10.1023/A:1023744019585)
 82. Buzdin A, Ustyugova S, Gogvadze E, **Lebedev Y**, Hunsmann G, Sverdlov E (2003). Genome-wide targeted search for human specific and polymorphic L1 integrations. *Hum Genet* 112 (56), 527–533, [10.1007/s00439-](https://doi.org/10.1007/s00439-)

83. Buzdin A, Ustyugova S, Khodosevich K, Mamedov I, **Lebedev Y**, Hunsmann G, Sverdlov E (2003). Human-specific subfamilies of HERV-K (HML-2) long terminal repeats: Three master genes were active simultaneously during branching of hominoid lineages. *Genomics* 81 (2), 149–156, [10.1016/S0888-7543\(02\)00027-7](#)
84. Buzdin AA, **Lebedev YB**, Sverdlov ED (2003). Human-Specific HERV-K Intron LTRs Have Nonaccidental Opposite Orientation Relative to the Direction of Gene Transcription and Might Be Involved in the Antisense Regulation of Gene Expression. *Russ. J. Bioorganic Chem.* 29 (1), 91–93, [10.1023/A:1022294906202](#)
85. Domansky AN, Akopov SB, **Lebedev YB**, Nikolaev LG, Sverdlov ED (2002). Enhancer activity of solitary long terminal repeat of human endogenous retrovirus K. *Russ. J. Bioorganic Chem.* 28 (4), 308–311, [10.1023/A:1019543808495](#)
86. Khodosevich K, **Lebedev Y**, Sverdlov E (2002). Endogenous retroviruses and human evolution. *Comp Funct Genomics* 3 (6), 494–498, [10.1002/cfg.216](#)
87. **Lebedev YB**, Bolorma B, Kzhyskowska JG, Ostashkin AS, Ilyin KV, Myandina GI, Pyagai PE, Itkes AV (2002). Integration of the type D Mason-Pfizer monkey virus into the human chromosome. *Mol Biol* 36 (6), 821–822, [10.1023/A:1021681909390](#)
88. Mamedov I, Batrak A, Buzdin A, Arzumanyan E, **Lebedev Y**, Sverdlov ED (2002). Genome-wide comparison of differences in the integration sites of interspersed repeats between closely related genomes. *Nucleic Acids Res* 30 (14), e71, [10.1093/nar/gnf071](#)
89. Buzdin A, Ustyugova S, Gogvadze E, Vinogradova T, **Lebedev Y**, Sverdlov E (2002). A new family of chimeric retrotranscripts formed by a full copy of U6 small nuclear RNA fused to the 3' terminus of L1. *Genomics* 80 (4), 402–406, [10.1006/geno.2002.6843](#)
90. Domansky AN, Akopov SB, **Lebedev YB**, Nikolaev LG, Sverdlov ED (2002). Enhancer activity of solitary long terminal repeat of human endogenous retrovirus K. *Bioorg Khim* 28 (4), 345.
91. Buzdin A, Khodosevich K, Mamedov I, Vinogradova T, **Lebedev Y**, Hunsmann G, Sverdlov E (2002). A technique for genome-wide identification of differences in the interspersed repeats integrations between closely related genomes and its application to detection of human-specific integrations of HERV-K LTRs. *Genomics* 79 (3), 413–422, [10.1006/geno.2002.6705](#)
92. Nadezhdin EV, **Lebedev YB**, Glazkova DV, Bornholdt D, Arman IP, Grzeschik KH, Hunsmann G, Sverdlov ED (2001). Identification of paralogous HERV-K LTRs on human chromosomes 3, 4, 7 and 11 in regions containing clusters of olfactory receptor genes. *Mol Genet Genomics* 265 (5), 820–825, [10.1007/s004380100476](#)
93. Kurdyukov SG, **Lebedev YB**, Artamonova II, Gorodentseva TN, Batrak AV, Mamedov IZ, Azhikina TL, Legchilina SP, Efimenko IG, Gardiner K, Sverdlov ED (2001). Full-sized HERV-K (HML-2) human endogenous retroviral LTR sequences on human chromosome 21: Map locations and evolutionary history. *Gene* 273 (1), 51–61, [10.1016/S0378-1119\(01\)00570-4](#)
94. Domansky AN, Kopantzev EP, Snezhkov EV, **Lebedev YB**, Leib-Mosch C, Sverdlov ED (2000). Solitary HERV-K LTRs possess bi-directional promoter activity and contain a negative regulatory element in the U5 region. *FEBS Lett* 472 (23), 191–195, [10.1016/S0014-5793\(00\)01460-5](#)
95. **Lebedev YB**, Belonovitch OS, Zybrova NV, Khil PP, Kurdyukov SG, Vinogradova TV, Hunsmann G, Sverdlov ED (2000). Differences in HERV-K LTR insertions in orthologous loci of humans and great apes. *Gene* 247 (12), 265–277, [10.1016/S0378-1119\(00\)00062-7](#)
96. **Lebedev YB** (2000). Endogenous retroviruses: A possible role in human cell function. *Mol Biol* 34 (4), 544–553, [10.1007/BF02759563](#)
97. Bogush ML, Velikodvorskaya TV, **Lebedev YB**, Nikolaev LG, Lukyanov SA, Fradkov AF, Pliyev BK, Boichenko MN, Usatova GN, Vorobiev AA, Andersen GL, Sverdlov ED (1999). Identification and localization of differences between *Escherichia coli* and *Salmonella typhimurium* genomes by suppressive subtractive hybridization. *Mol Gen Genet* 262 (45), 721–729, [10.1007/s004380051134](#)
98. Belyaeva OV, Balanovsky OP, Ashworth LK, **Lebedev YB**, Spitsyn VA, Guseva NA, Erdes S, Mikulich AI, Khusnutdinova EK, Limborska SA (1999). Fine mapping of a polymorphic CA repeat marker on human chromosome 19 and its use in population studies. *Gene* 230 (2), 259–266, [10.1016/S0378-1119\(99\)00056-6](#)
99. Lavrentieva I, Broude NE, **Lebedev Y**, Gottesman II, Lukyanov SA, Smith CL, Sverdlov ED (1999). High

- polymorphism level of genomic sequences flanking insertion sites of human endogenous retroviral long terminal repeats. *FEBS Lett* 443 (3), 341–347, [10.1016/S0014-5793\(99\)00004-6](https://doi.org/10.1016/S0014-5793(99)00004-6)
100. Lapuk AV, Khil PP, Lavrentieva IV, **Lebedev YB**, Sverdlov ED (1999). A human endogenous retrovirus-like (HERV) LTR formed more than 10 million years ago due to an insertion of HERV-H LTR into the 5' LTR of HERV-K is situated on human chromosomes 10, 19 and Y. *J Gen Virol* 80 (4), 835–839, [10.1099/0022-1317-80-4-835](https://doi.org/10.1099/0022-1317-80-4-835)
 101. Khil PP, **Lebedev YB**, Sverdlov ED (1998). A new putative gene preferentially expressed in the human brain located on chromosome 19q12 near the human endogenous virus HERV-K LTR. *Russ. J. Bioorganic Chem.* 24 (1), 62–63.
 102. Khil PP, **Lebedev YB**, Sverdlov ED (1998). The human endogenous virus HERV-K long terminal repeat in an intron of the ZNF91 gene. *Russ. J. Bioorganic Chem.* 24 (2), 112–116.
 103. Khil PP, **Lebedev YB**, Sverdlov ED (1998). A New Putative Gene Preferentially Expressed in the Human Brain Located on Chromosome 19q12 near the Human Endogenous Virus HERV-K LTR. *Bioorg Khim* 24 (1), 73–74.
 104. Klinov DV, Lagutina IV, Prokhorov VV, Neretina T, Khil PP, **Lebedev YB**, Cherny DI, Demin VV, Sverdlov ED (1998). High resolution mapping DNAs by R-loop atomic force microscopy. *Nucleic Acids Res* 26 (20), 4603–4610, [10.1093/nar/26.20.4603](https://doi.org/10.1093/nar/26.20.4603)
 105. Lavrentieva I, Khil P, Vinogradova T, Akhmedov A, Lapuk A, Shakhova O, **Lebedev Y**, Monastyrskaya G, Sverdlov ED (1998). Subfamilies and nearest-neighbour dendrogram for the LTRs of human endogenous retroviruses HERV-K mapped on human chromosome 19: Physical neighbourhood does not correlate with identity level. *Hum Genet* 102 (1), 107–116, [10.1007/s004390050662](https://doi.org/10.1007/s004390050662)
 106. Akopov SB, Nikolaev LG, Khil PP, **Lebedev YB**, Sverdlov ED (1998). Long terminal repeats of human endogenous retrovirus K family (HERV-K) specifically bind host cell nuclear proteins. *FEBS Lett* 421 (3), 229–233, [10.1016/S0014-5793\(97\)01569-X](https://doi.org/10.1016/S0014-5793(97)01569-X)
 107. Khil PP, **Lebedev YB**, Sverdlov ED (1998). The Human Endogenous Virus HERV-K Long Terminal Repeat in an Intron of the ZNF91 Gene. *Bioorg Khim* 24 (2), 130–131.
 108. Khil PP, Kostina MB, Azhikina TL, Kolesnik TB, **Lebedev YB**, Sverdlov ED (1997). Structural Features of Four Long Terminal Repeats of Human Endogenous Retroviruses and Their Integration Sites. *Bioorg Khim* 23 (5), 440.
 109. Vinogradova T, Volik S, **Lebedev Y**, Shevchenko Y, Lavrentyeva I, Khil P, Grzeschik KH, Ashworth LK, Sverdlov E (1997). Positioning of 72 potentially full size LTRs of human endogenous retroviruses HERV-K on the human chromosome 19 map. Occurrences of the LTRs in human gene sites. *Gene* 199 (12), 255–264, [10.1016/S0378-1119\(97\)00376-4](https://doi.org/10.1016/S0378-1119(97)00376-4)
 110. Khil PP, Kostina MB, Azhikina TL, Kolesnik TB, **Lebedev YB**, Sverdlov ED (1997). Structural features of four long terminal repeats of human endogenous retroviruses and their integration sites. *Russ. J. Bioorganic Chem.* 23 (5), 406–411.
 111. **Lebedev Y**, Akopyants N, Azhikina T, Shevchenko Y, Potapov V, Stecenko D, Berg D, Sverdlov E (1996). Oligonucleotides containing 2-aminoadenine and 5-methylcytosine are more effective as primers for PCR amplification than their nonmodified counterparts. *Genet Anal Tech Appl* 13 (1), 15–21, [10.1016/1050-3862\(96\)00139-8](https://doi.org/10.1016/1050-3862(96)00139-8)
 112. **Lebedev YB**, Volik SV, Obradovic D, Ermolaeva OD, Ashworth LK, Lennon GG, Sverdlov ED (1995). Physical mapping of sequences homologous to an endogenous retrovirus LTR on human chromosome 19. *Mol Gen Genet* 247 (6), 742–748, [10.1007/BF00290406](https://doi.org/10.1007/BF00290406)
 113. Volik S, **Lebedev Y**, Nikolaev L, Shevchenko Y, Vinogradova T, Kopantzev E, Kolesnik T, Monastyrskaya G, Kunz U, Grzeschik KH, Ashworth LK, Lennon G, Sverdlov E (1995). Mapping of transcribed sequences on human chromosome 19. *Mitochondrial DNA* 6 (1), 13–26, [10.3109/10425179509074694](https://doi.org/10.3109/10425179509074694)
 114. Borodin A, Kopantzev E, Wagner L, Volik S, Ermolaeva O, **Lebedev Y**, Monastyrskaya G, Kunz J, Grzeschik KH, Sverdlov E (1995). An arrayed library enriched in hncDNA corresponding to transcribed sequences of human chromosome 19: preparation and analysis. *Genet Anal Tech Appl* 12 (1), 23–31, [10.1016/1050-3862\(95\)00106-9](https://doi.org/10.1016/1050-3862(95)00106-9)