

Резюме: Лебедев Юрий Борисович

Адрес

Федеральное государственное бюджетное учреждение науки Институт биоорганической химии им. академиков М.М. Шемякина и Ю.А. Овчинникова Российской академии наук, Москва, Россия

Контакты

<https://www.ibch.ru/ru/users/25>

Образование

1973–1978

Москва

МГУ

Работа в ИБХ

2018–наст.вр.

Главный научный сотрудник

Членство в советах и комиссиях ИБХ

Диссертационный совет

Ученый совет

Научные интересы

Руководитель лаборатории доктор биологических наук Юрий Борисович Лебедев является одним из ведущих отечественных специалистов в области изучения структуры, функционирования и эволюции генома млекопитающих. Исследования в этой области начаты Ю. Б. Лебедевым в начале 1990-х, когда он с группой молодых ученых лаборатории академика Е. Д. Свердлова вошли в число первых участников Российской программы «Геном Человека». Как руководитель лаборатории Ю. Б. Лебедев продолжает разрабатывать оригинальное направление эволюционной геномики, связанное с исследованием функциональных последствий распространения и активации ретропозонов в геномах высших приматов.

Членство в сообществах

В 1996 г. Лебедев Ю. Б. был принят в члены HUGO.

Степени и звания

Профессор

Доктор наук (Биологические науки, 03.00.03 — Молекулярная биология)

Гранты и проекты

2020–
2022 [Взаимодействие генотипа и репертуара Т-лимфоцитов при формировании противовирусного иммунного ответа](#)

2015–
2019 [Изучение динамики системы клеточного адаптивного иммунитета при флавивирусной инфекции](#)

2017–
2020 [Анализ клонального спектра малых субпопуляций Т-лимфоцитов при формировании иммунного ответа у человека](#)

Публикации

1. Salnikova MA, **Lebedev YB** (2024). Longitudinal tracking of T-cell repertoire reveals long-lasting CD4⁺ yellow fever specific clone cluster. *Russian Journal of Infection and Immunity* 14 (3), 539–543, [10.15789/2220-7619-LTO-16665](https://doi.org/10.15789/2220-7619-LTO-16665)
2. Smirnova AO, Miroshnichenkova AM, Belyaeva LD, Kelmanson IV, **Lebedev YB**, Mamedov IZ, Chudakov DM, Komkov AY (2023). Novel bimodal TRBD1-TRBD2 rearrangements with dual or absent D-region contribute to TRB V-(D)-J combinatorial diversity. *Front Immunol* 14, 1245175, [10.3389/fimmu.2023.1245175](https://doi.org/10.3389/fimmu.2023.1245175)
3. Smirnova AO, Miroshnichenkova AM, Olshanskaya YV, Maschan MA, **Lebedev YB**, Chudakov DM, Mamedov IZ, Komkov A (2023). The use of non-functional clonotypes as a natural calibrator for quantitative bias correction in adaptive immune receptor repertoire profiling. *Elife* 12, , [10.7554/eLife.69157](https://doi.org/10.7554/eLife.69157)
4. Komech EA, Koltakova AD, Barinova AA, Minervina AA, Salnikova MA, Shmidt EI, Korotaeva TV, Loginova EY, Erdes SF, Bogdanova EA, Shugay M, Lukyanov S, **Lebedev YB**, Zvyagin IV (2022). TCR repertoire profiling revealed antigen-driven CD8⁺ T cell clonal groups shared in synovial fluid of patients with spondyloarthritis. *Front Immunol* 13, 973243, [10.3389/fimmu.2022.973243](https://doi.org/10.3389/fimmu.2022.973243)
5. Sycheva AL, Komech EA, Pogorelyy MV, Minervina AA, Urazbakhtin SZ, Salnikova MA, Vorovitch MF, Kopantzev EP, Zvyagin IV, Komkov AY, Mamedov IZ, **Lebedev YB** (2022). Inactivated tick-borne encephalitis vaccine elicits several overlapping waves of T cell response. *Front Immunol* 13, 970285, [10.3389/fimmu.2022.970285](https://doi.org/10.3389/fimmu.2022.970285)
6. Urazbakhtin S, Smirnova A, Volakhava A, Zerkalnikova E, Salyutina M, Doubek M, Jelinkova H, Khudainazarova N, Volchikov E, Belyaeva L, Komech E, Pavlova S, **Lebedev Y**, Plevova K, Olshanskaya Y, Komkov A, Mamedov I (2022). The Absence of Retroelement Activity Is Characteristic for Childhood Acute Leukemias and Adult Acute Lymphoblastic Leukemia. *Int J Mol Sci* 23 (3), , [10.3390/ijms23031756](https://doi.org/10.3390/ijms23031756)
7. **(конференция)** Sycheva AL, Pogorelyy MV, Komech EA, Urazbakhtin SZ, Minervina AA, Kopancev EP, Vorovitch MF, Zvyagin IV, Mamedov IZ, **Lebedev YB** (2021). Features of T-cell immune response to tick-borne encephalitis vaccine. *Eur J Immunol* 51 (S1), 1–448, <https://doi.org/10.1002/eji.202170200>
8. **(конференция)** Zvyagin IV, Blagov S, Fomchenkova V, Fadeeva M, Komech EA, Zhogov V, Barinova AA, Mikelov AI, Sycheva AL, **Lebedev YB**, Maschan MA (2021). T cell repertoire sequencing to study the contribution of different donor T cell subsets to patient repertoire at the early stage after αβT/CD19-depleted allogeneic hematopoietic stem cell transplantation. *Eur J Immunol* 51 (S1), 1–448, <https://doi.org/10.1002/eji.202170200>
9. **(конференция)** Комеч ЕА, Звягин ИВ, **Лебедев ЮБ**, Сальникова МА, , Минервина АА (2021). T-CELL REPERTOIRE OF SYNOVIAL FLUID IN SPONDYLOARTHROPATHIES EXHIBITS HALLMARKS OF HLA-DEPENDENT CLONAL EXPANSIONS AND REMAINS STABLE OVER 1.5 YEARS. *Ann Rheum Dis* (80), 204, [10.1136/annrheumdis-2021-eular.3498](https://doi.org/10.1136/annrheumdis-2021-eular.3498)
10. Kovalenko EI, Zvyagin IV, Streltsova MA, Mikelov AI, Erokhina SA, Telford G, Sapozhnikov AM, **Lebedev YB** (2021). Surface NKG2C identifies differentiated αβT-cell clones expanded in peripheral blood. *Front Immunol* 11, 613882, [10.3389/fimmu.2020.613882](https://doi.org/10.3389/fimmu.2020.613882)
11. Minervina AA, Komech EA, Titov A, Koraichi MB, Rosati E, Mamedov IZ, Franke A, Efimov GA, Chudakov DM, Mora T, Walczak AM, **Lebedev YB**, Pogorelyy MV (2021). Longitudinal high-throughput TCR repertoire profiling reveals the dynamics of T-cell memory formation after mild COVID-19 infection. *Elife* 10, 1–17, [10.7554/eLife.63502](https://doi.org/10.7554/eLife.63502)
12. Komkov AY, Urazbakhtin SZ, Saliutina MV, Komech EA, Shelygin YA, Nugmanov GA, Shubin VP, Smirnova AO, Bobrov MY, Tsukanov AS, Snezhkina AV, Kudryavtseva AV, **Lebedev YB**, Mamedov IZ (2020). SeqURE – a new copy-capture based method for sequencing of unknown Retroposition events. *Mob DNA* 11 (1), 33, [10.1186/s13100-020-00228-6](https://doi.org/10.1186/s13100-020-00228-6)
13. **(конференция)** Mikelov AI, Komech EA, **Lebedev YB**, Zvyagin IV (2020). In- and off- season peripheral blood T cell repertoire profiling of patients with birch pollen allergy. *Allergy* 75 (S109), 188, [10.1111/all.14506](https://doi.org/10.1111/all.14506)
14. **(конференция)** Комков АЮ, Мамедов ИЗ, **Лебедев ЮБ**, Атапина Е (2020). A cost-effective quasi single-cell assay for deciphering of clonal architecture of leukemic cells. *Klin Padiatr* 232 (3), e7, [10.1055/s-0040-1709799](https://doi.org/10.1055/s-0040-1709799)
15. Minervina AA, Pogorelyy MV, Komech EA, Karnaukhov VK, Bacher P, Rosati E, Franke A, Chudakov D, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2020). Primary and secondary anti-viral response captured by the dynamics and phenotype of individual T cell clones. *Elife* 9, , [10.7554/eLife.53704](https://doi.org/10.7554/eLife.53704)

16. Rosati E, Pogorelyy MV, Dowds CM, Moller FT, Sorensen SB, **Lebedev YB**, Frey N, Schreiber S, Spehlmann ME, Andersen V, Mamedov IZ, Franke A (2019). Identification of disease-associated traits and clonotypes in the T-cell receptor repertoire of monozygotic twins affected by inflammatory bowel diseases. *J Crohns Colitis* 14 (6), 778–790, [10.1093/ecco-jcc/jjz179](https://doi.org/10.1093/ecco-jcc/jjz179)
17. Микелов АИ, Староверов ДБ, Комеч ЕА, **Лебедев ЮБ**, Чудаков ДМ, Zvyagin IV (2019). Correlated dynamics of serum IGE and IGE+ clonotype count with allergen air level in seasonal allergic rhinitis. *Bulletin of Russian State Medical University* 5 (5), 13–22, [10.24075/brsmu.2019.072](https://doi.org/10.24075/brsmu.2019.072)
18. Komkov A, Miroshnichenkova A, Nugmanov G, Popov A, Pogorelyy M, Zapletalova E, Jelinkova H, Pospisilova S, **Lebedev Y**, Chudakov D, Olshanskaya Y, Plevova K, Maschan M, Mamedov I (2019). High-throughput sequencing of T-cell receptor alpha chain clonal rearrangements at the DNA level in lymphoid malignancies. *Br J Haematol* 188 (5), 723–731, [10.1111/bjh.16230](https://doi.org/10.1111/bjh.16230)
19. **(конференция)** Mikelov AI, Turchaninova MA, Komech EA, Staroverov DB, Shvets SM, **Lebedev YB**, Chudakov DM, Zvyagin IV (2019). Longitudinal profiling of immunoglobulin heavy-chain repertoires in memory B-cells, plasmablasts and plasma cells from peripheral blood of individuals with birch pollen allergy. *Allergy* 74 (S106), 174.
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21. Nugmanov GA, Komkov AY, Saliutina MV, Minervina AA, **Lebedev YB**, Mamedov IZ (2019). [A Pipeline for the Error-free Identification of Somatic Alu Insertions in High-throughput Sequencing Data]. *Mol Biol (Mosk)* 53 (1), 154–165, [10.1134/S0026898419010117](https://doi.org/10.1134/S0026898419010117)
22. Nugmanov GA, Komkov AY, Saliutina MV, Minervina AA, **Lebedev YB**, Mamedov IZ (2019). A Pipeline for the Error-Free Identification of Somatic Alu Insertions in High-Throughput Sequencing Data. *Mol Biol* 53 (1), 138–146, [10.1134/S0026893319010114](https://doi.org/10.1134/S0026893319010114)
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24. Komkov AY, Minervina AA, Nugmanov GA, Saliutina MV, **Lebedev YB**, Mamedov IZ, Khodosevich KV (2018). An advanced enrichment method for rare somatic retroelement insertions sequencing. *Mob DNA* 9 (1), 31, [10.1186/s13100-018-0136-1](https://doi.org/10.1186/s13100-018-0136-1)
25. **(конференция)** Комеч ЕА, Колтакова АД, Мясоутова АА, Коротаева ТВ, Шмидт НИ, Шостак НА, **Лебедев ЮБ**, Звягин ИВ (2018). Клональная характеристика Т-лимфоцитов очага воспаления у больных со спондилоартропатиями. *Nauchno-Prakticheskaya Revmatologiya* 56 (3), 91.
26. **(конференция)** Звягин ИВ, **Лебедев ЮБ**, Мясоутова АА, Комеч ЕА (2018). TCRbeta CDR3 motif is detected in synovial fluid of patients with different spondyloarthropathies. *FEBS Open Bio* 8, 489.
27. **(конференция)** Fomchenkova E, Komech A, Blagov , Sycheva L, **Lebedev B**, Chudakov M, Maschan A, Zvyagin V (2018). T cell repertoire profiling after hematopoietic stem cell transplantation with CD19/αβT cell depletion and donor lymphocyte infusion. *FEBS Open Bio* 8 (S1), 281: P.09–230–Tue.
28. Komech EA, Pogorelyy MV, Egorov ES, Britanova OV, Rebrikov DV, Bochkova AG, Shmidt EI, Shostak NA, Shugay M, Lukyanov S, Mamedov IZ, **Lebedev YB**, Chudakov DM, Zvyagin IV (2018). CD8+T cells with characteristic T cell receptor beta motif are detected in blood and expanded in synovial fluid of ankylosing spondylitis patients. *Rheumatology (Oxford)* 57 (6), 1097–1104, [10.1093/rheumatology/kex517](https://doi.org/10.1093/rheumatology/kex517)
29. Komech EA, Zvyagin IV, Pogorelyy MV, Mamedov IZ, Fedorenko DA, **Lebedev YB** (2018). Characterization of the T-cell repertoire after autologous HSCT in patients with ankylosing spondylitis. *Acta Naturae* 10 (2), 48–57, [10.32607/2075851-2018-10-2-48-57](https://doi.org/10.32607/2075851-2018-10-2-48-57)
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31. Pogorelyy MV, Minervina AA, Chudakov DM, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2018). Method for identification of condition-associated public antigen receptor sequences. *Elife* 7, , [10.7554/eLife.33050](https://doi.org/10.7554/eLife.33050)

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33. Komech EA, Zvyagin IV, Pogorelyy MV, Mamedov IZ, Fedorenko DA, **Lebedev YB** (2018). Characterization of the T-cell Repertoire after Autologous HSCT in Patients with Ankylosing Spondylitis. *Acta Naturae* 10 (2), 48–57, [10.32607/20758251-2018-10-2-48-57](https://doi.org/10.32607/20758251-2018-10-2-48-57)
34. Pogorelyy MV, Elhanati Y, Marcou Q, Sycheva AL, Komech EA, Nazarov VI, Britanova OV, Chudakov DM, Mamedov IZ, **Lebedev YB**, Mora T, Walczak AM (2017). Persisting fetal clonotypes influence the structure and overlap of adult human T cell receptor repertoires. *PLoS Comput Biol* 13 (7), e1005572, [10.1371/journal.pcbi.1005572](https://doi.org/10.1371/journal.pcbi.1005572)
35. Komkov A, Miroshnichenkova A, Minervina A, Nugmanov G, **Lebedev Y**, Mamedov I, Olshanskaya Y, Maschan M (2017). High-throughput sequencing for diagnostics of minimal residual disease in acute lymphoblastic leukemia. *Klin Padiatr* , , [10.1055/s-0037-1602224](https://doi.org/10.1055/s-0037-1602224)
36. (конференция) Pogorelyy M, PuelmaTouzer M, Minervina AA, Sycheva AL, Chudakov DM, Mamedov IZ, Mora T, Walczak AM, **Lebedev YB** (2017). High throughput sequencing of identical twins TCR repertoires after yellow fever vaccination. , 60.
37. Zvyagin IV, Mamedov IZ, Tatarinova OV, Komech EA, Kurnikova EE, Boyakova EV, Brilliantova V, Shelikhova LN, Balashov DN, Shugay M, Sycheva AL, Kasatskaya SA, **Lebedev YB**, Maschan AA, Maschan MA, Chudakov DM (2017). Tracking T-cell immune reconstitution after TCR $\alpha\beta$ /CD19-depleted hematopoietic cells transplantation in children. *Leukemia* 31 (5), 1145–1153, [10.1038/leu.2016.321](https://doi.org/10.1038/leu.2016.321)
38. (конференция) Сычева АЛ, Погорелый МВ, Комеч ЕА, Звягин ИВ, Мамедов ИЗ, **Лебедев ЮБ** (2017). Изучение малых субпопуляций активированных Т-лимфоцитов из крови доноров, вакцинированных против вируса жёлтой лихорадки. , 98.
39. (конференция) Zvyagin I, Tatarinova O, Mamedov I, Komech E, Maschan A, Shelikhova L, Kurnikova E, Boyakova E, **Lebedev Y**, Maschan M, Chudakov D (2016). T Cell Repertoire after Alpha/Beta-T Cell Depleted Allogeneic Hematopoietic Stem Cell Transplantation in Pediatric Patients. *Blood* (128), 4582.
40. Nazarov VI, Minervina AA, Komkov AY, Pogorelyy MV, Maschan MA, Olshanskaya YV, Zvyagin IV, Chudakov DM, **Lebedev YB**, Mamedov IZ (2016). Reliability of immune receptor rearrangements as genetic markers for minimal residual disease monitoring. *Bone Marrow Transplant* 51 (10), 1408–1410, [10.1038/bmt.2016.148](https://doi.org/10.1038/bmt.2016.148)
41. (конференция) Komkov AY, Minervina AA, Pogorelyy MV, Zvyagin IV, Panferova A, Olshanskaya Y, Chudakov DM, Maschan M, Mamedov IZ, **Lebedev YB** (2016). Next generation sequencing based approach for monitoring of minimal residual disease in acute lymphoblastic leukemia. *FEBS J* 283 (S1), 376, [10.1111/febs.13808](https://doi.org/10.1111/febs.13808)
42. (конференция) Погорелый МВ, Сычева АЛ, Мамедов ИЗ, Мора Т, Вальзак АМ, **Лебедев ЮБ** (2016). Клоны Т-клеток пуповинной крови обнаруживаются в репертуарах Т-клеточных рецепторов взрослых доноров. , 23.
43. Minervina AA, Komkov AY, Mamedov IZ, **Lebedev YB** (2016). Advanced lymphoblastic clones detection in T-cell leukemia. *Dokl Biochem Biophys* 467 (1), 85–88, [10.1134/S1607672916020022](https://doi.org/10.1134/S1607672916020022)
44. (конференция) Сычева АЛ, Погорелый МВ, Комеч ЕА, Мамедов ИЗ, **Лебедев ЮБ** (2016). Динамика Т-клеточного репертуара человека в ходе противогриппозной вакцинации и ревакцинации. , 27.
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46. Nazarov VI, Pogorelyy MV, Komech EA, Zvyagin IV, Bolotin DA, Shugay M, Chudakov DM, **Lebedev YB**, Mamedov IZ (2015). tcR: An R package for T cell receptor repertoire advanced data analysis. *BMC Bioinformatics* 16 (1), 175, [10.1186/s12859-015-0613-1](https://doi.org/10.1186/s12859-015-0613-1)
47. (конференция) Pogorelyy MV, Sycheva AL, Komech EA, Marcou Q, Elhanati Y, Mora T, Walczak A, Mamedov IZ, **Lebedev YB** (2015). Deep TCR repertoire profiling after seasonal influenza vaccination. , 432.
48. Kurnosov AA, Ustyugova SV, Nazarov VI, Minervina AA, Komkov AY, Shugay M, Pogorelyy MV, Khodosevich KV, Mamedov IZ, **Lebedev YB** (2015). The evidence for increased L1 activity in the site of

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 54. Kurnosov AA, Ustyugova SV, Pogorelyy MV, Komkov AY, Bolotin DA, Khodosevich KV, Mamedov IZ, **Lebedev YB** (2013). A novel approach to identification of somatic retroelements' insertions in human genome. *Russ. J. Bioorganic Chem.* 39 (4), 417–425, [10.1134/S1068162013040110](https://doi.org/10.1134/S1068162013040110)
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